(DP321) DNAquick Plant System -Plant

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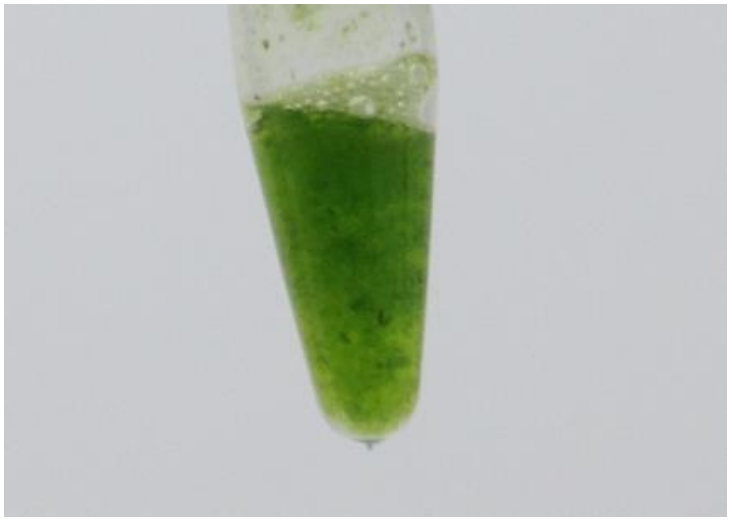
Ver. No. 20170328

Experiment Preparation

1. Plant Leaves
2. Mortar; Liquid nitrogen
3. Pipette and matched sterile tips (10 μl, 200 μl, 1 ml); 1.5 ml centrifuge tubes
4. Isopropanol; 70% ethanol; Clean absorbent paper
5. Vortex oscillator; Dry bath/water bath; Centrifuge

Step 1

Add liquid nitrogen to 100 mg fresh plant tissue or 20 mg dry weight tissue, and fully grind the tissues.



Add 400 μl Buffer FP1 and 6 μl RNase A (10 mg/ml). After vortex for 1 min, place at room temperature for 10 min.

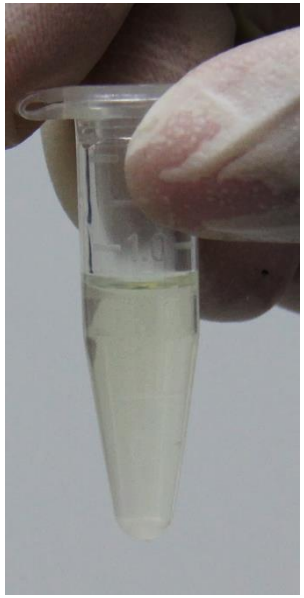
Step 2

Add 130 μl Buffer FP2, mix well by vortex for 1min.

Step 3

Centrifuge at 12,000 rpm (~13,400 g) for 5 min, and transfer the supernatant to a new centrifuge tube.

Step 4



Add 0.7 × volume of isopropanol to the supernatant and mix well. There may be flocculent genome DNA at this time. (for example, add 350 μl isopropanol to 500 μl supernatant),

Centrifuge at 12,000 rpm (~13,400 g) for 2 min, discard supernatant and retain precipitation.

Step 5

Add 600 μl buffer 70% ethanol, and mix well by vortex for 5 sec.

Centrifuge at 12,000 rpm (~13,400 g) for 2 min, and discard the supernatant.

Step 6 重复操作步骤5。

Step 6

Open cover and place on clean absorbent paper at room temperature for 5-10 minutes to completely dry the ethanol.

Note: Ethanol residues can inhibit subsequent enzymatic reactions (restriction enzyme digestion, PCR, etc.) experiments.

Step 7

Add appropriate amount of Buffer TE, and incubate at 65℃ for 10-60 min to dissolve DNA. Mix well upside down for several times during the period.