

# (DP348) TIANamp Blood DNA Kit -200 µl~1 ml Anticoagulant whole blood

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Ver. No. 20170328

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# **Experiment Preparation**

- 1. Anticoagulant blood 200 µl~1 ml
- 2. Pipette and matched sterile tips (200 µl, 1 ml)
- 3. 96-100% ethanol
- 4. Vortex oscillator; Dry bath/water bath; Centrifuge





Note: This experiment takes human blood as an example. The whole blood of mammals can be extracted by this process. For anticoagulant blood of poultry, birds, amphibians or lower organisms, the red blood cells are nucleated cells, so the starting sample volume is 5-20 µl, and Buffer GS should be added to top up to 200 µl. For blood clot samples, the Liquefaction Columns CX1 (TIANGEN, RK165) (self-provided) can be selected to treat the blood clot.



#### **Experiment Preparation - Kit Preparation**

Please add 96-100% ethanol in Buffer PWB before use according to the volume indicated on the label of the bottle.







Add 1-2.5  $\times$  volume of Buffer CL in 1 ml whole blood, and mix well. Centrifuge at 10,000 rpm (~11,500 × g ) for 1 min.

Remove supernatant, leave leukocyte precipitate, and add 200 µl Buffer GS and vortex until thoroughly mixed.





Add 200 µl Buffer GB Mix well upside and 20 µl Proteinase K. down.

Incubate at 56°C for 10 min.

The solution should become clear at this step.





Incubate at room temperature for 2-5 min, then add 350 µl Buffer BD



Mix well upside down, and there may be flocculent precipitate at this step.











Place Spin Column CG2 in a collection tube. Transfer the solution and flocculent precipitate from the previous step to Spin Column CG2.

Centrifuge at 12,000 rpm  $(~13,400 \times g)$  for 30 sec. Discard the waste liquid in the collection tube and place the Spin Column CG2 back in the collection tube.







Add 500 µl Buffer GDB to Spin Column CG2. Centrifuge at 12,000 rpm ( $\sim$ 13,400×g) for 30 sec, discard the waste liquid in the collection tube and place the Spin Column CG2 into the collection tube.







Add 600 µl Buffer PWB to Spin Column CG2 (ensure 96-100% ethanol has been added before use).

Centrifuge at 12,000 rpm ( $\sim$ 13,400 $\times$ g) for 30 sec, discard waste liquid in the collection tube and place the Spin Column CG2 into the collection tube.

#### Step 7 Repeat step 6.





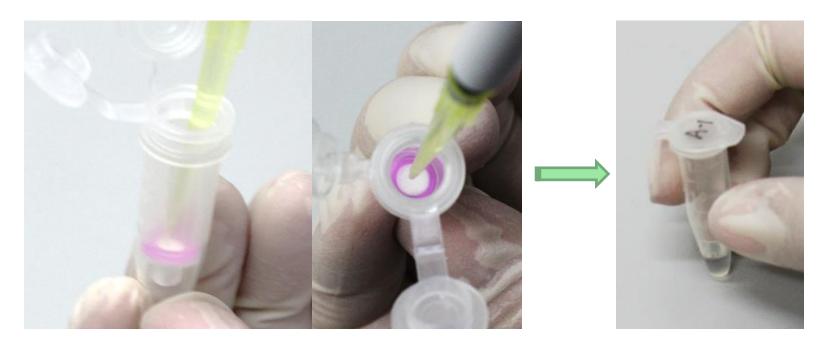


Centrifuge at 12,000 rpm ( $\sim$ 13,400 $\times$ g) for 2 min, and discard the waste liquid.

Place the Spin Column CG2 at room temperature for 2 minutes to completely dry Buffer PWB in the membrane.

Note: Ethanol residues in Buffer PWB can inhibit subsequent enzymatic reactions (restriction enzyme digestion, PCR, etc.) experiments.





Transfer the Spin Column CG2 into a 1.5 ml centrifuge tube, and add 50-200  $\mu$ l Buffer TB to the middle of the adsorption membrane. Place at room temperature for 2 min and centrifuge at 12,000 rpm (~13,400×g) for 2 min to collect the solution into the centrifuge tube.